

ON THE OCCURRENCE OF HAEMOGREGARINA NUCLEOBISECANS SHORTT, 1917 IN TOADS OF WEST BENGAL

R. RAY

Zoological Survey of India, Calcutta

ABSTRACT

Haemogregarina nucleobisecans was found to infect two species of toad viz., *Bufo melanostictus* (13.4%) and *B. andersoni* (28%) from Bankura and Purulia districts, West Bengal, India. Intra-erythrocytic development and tissue schizogony of the parasite in the liver parenchyma from both the bufonid hosts were described and illustrated. *H. nucleobisecans* has been recorded from a new geographical locale of Eastern India and from a new host *Bufo andersoni*.

INTRODUCTION

Shortt (1917) described *Haemogregarina nucleobisecans* for the first time from *Bufo melanostictus* collected from Cawnpore, Delhi and Ambala (Punjab), India. Subsequently, Wenyon (1926) reported the occurrence of this parasite from the same host in India. Since then there was no information on amphibian haemogregarines till the reports of Ray (1977, 1979). Sinha (1979) reported one *Haemogregarina* sp. from *Bufo himalayanus* of Darjeeling, West Bengal.

The present paper deals with the detailed description of *Haemogregarina nucleobisecans* in two species of Toad viz., *Bufo melanostictus* and *B. andersoni* along with the schizogony in the liver of both the hosts. *Bufo andersoni* has been recorded as a new host for this haemogregarine parasite.

MATERIAL AND METHODS

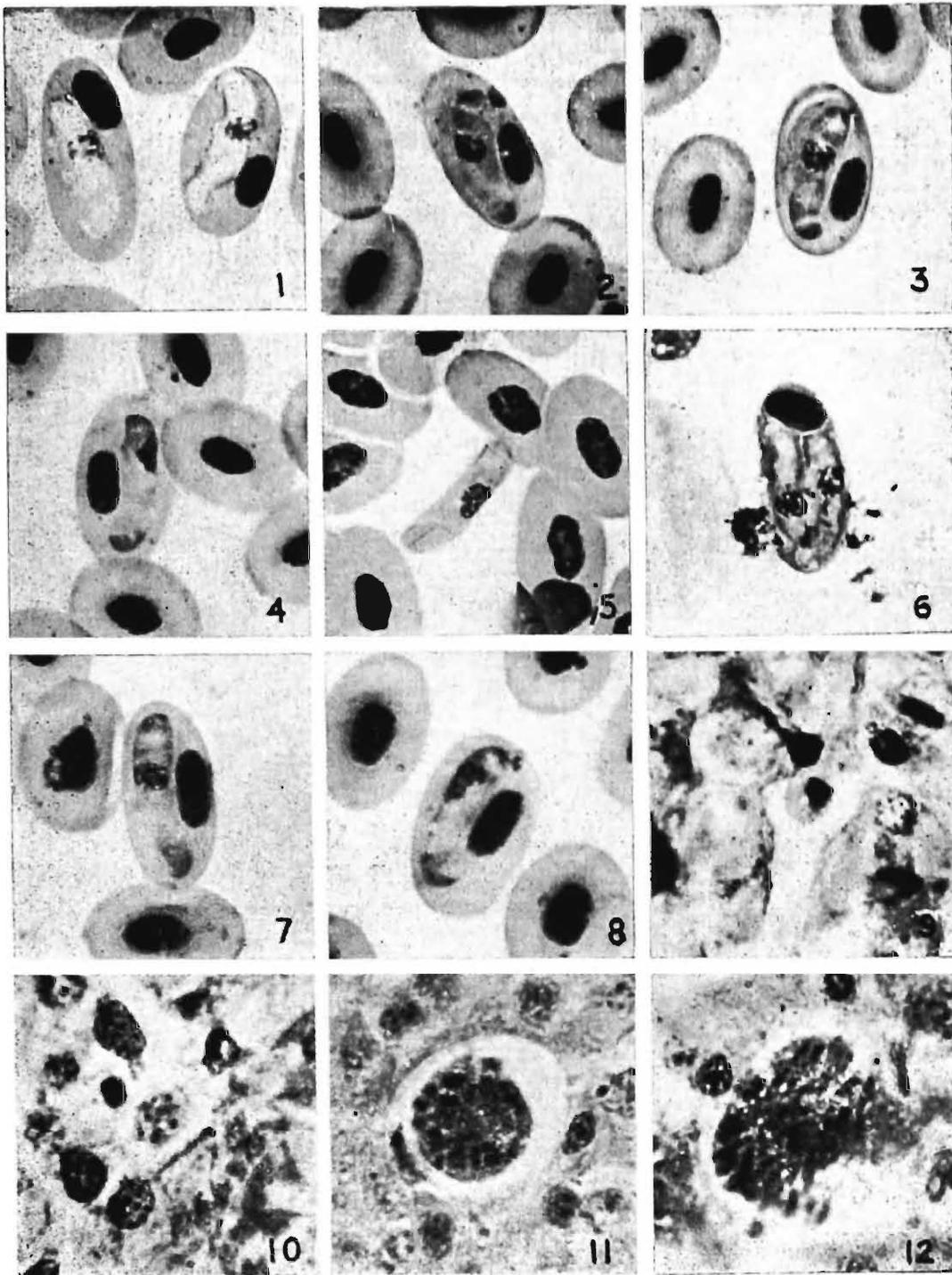
Both the hosts were collected from Bankura and Purulia districts of West Bengal and some of them were kept alive in the laboratory for examination. Peripheral blood was obtained from finger tips on alternate

days and at autopsy. Impression and spread preparations were made from liver, lungs, kidney etc. Air-dried blood films and organ imprints were fixed in 100% methanol and stained with Ramanowsky type of stains. For microtome section the tissue of lung, liver, kidney and spleen were fixed in alcoholic Bouin's fixative, followed by general histological technique (Pearse, 1960) and stained with iron-haematoxylin and eosin.

Measurements were obtained from the camera-lucida drawings drawn on a graph paper (mm division) as it facilitated the area measurements by counting the squares covered. The photomicrographs were taken with the help of 'Ergavel C. Z. microscope using PM6 attachment camera.

OBSERVATION

47 (13.4%) out of 343 examples of *Bufo melanostictus* and 7 (28%) out of 25 *B. andersoni* examined were found positive for the parasite. A minimum of 5% to a maximum of 15% of the total R. B. C. counted were found to be infected with *H. nucleobisecans*. The description of the parasite is as follows :



Figs. 1-12. Photomicrographs of the erythrocytic and tissue stages of *Haemogregarina nucleobisecans* from *Bufo melanostictus* and *Bufo andersoni*.

- | | |
|---|--------|
| 1-4. Elongate mature gametocytes in the peripheral blood of <i>B. melanostictus</i> . | X 1665 |
| 5. An extra-copular free gametocyte | X 1800 |
| 6. An erythrocyte with double infection | X 1665 |
| 7-8. Elongate mature gametocytes in <i>Bufo andersoni</i> . | X 1800 |
| 9. A mononucleated schizont within the liver tissue. | X 1110 |
| 10. An immature schizont | X 1400 |
| 11. A multinucleated mature schizont within the liver parenchyma. | X 1140 |
| 12. A mature schizont with developing merozoites. | X 1665 |

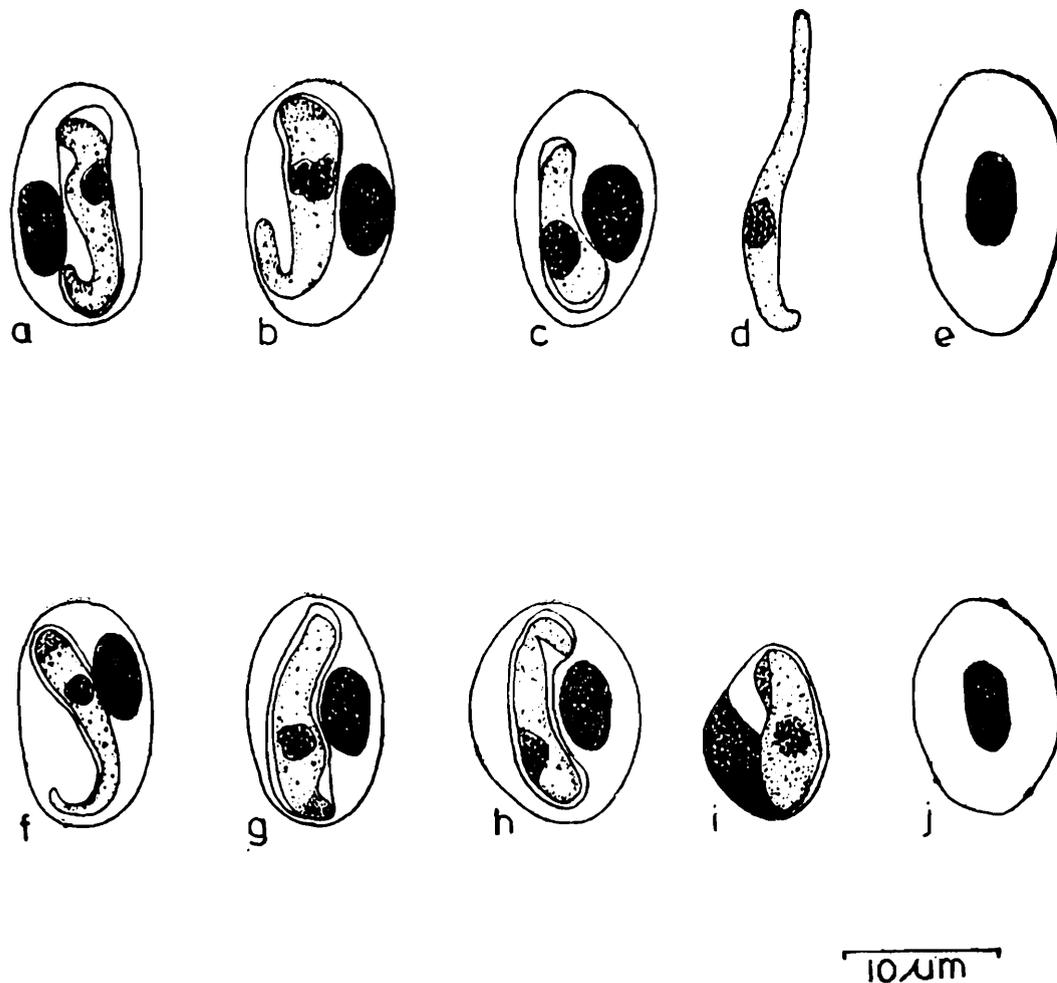


Fig. 1. Camera-lucida drawings of the erythrocytic stages of *Haemogregarina nucleobisecans* from two anuran hosts.

a-d, the gametocyte stages of *H. nucleobisecans* in *Bufo melanostictus*. a & b, the elongate mature forms ; c, the young forms ; d, the free gametocyte ; e, the uninfected erythrocyte ; f-i, the elongate mature gametocyte stages of *H. nucleobisecans* in *Bufo andersoni* ; j, the uninfected erythrocyte.

Gametocytes :

Elongate mature (Figs. 1a-c & Pl. V Figs. 1-4) :

N=15. These are sausage shaped endoparasites of erythrocytes, slightly curved and lying with its concave border applied to the convexity of the host cell nucleus or *visè-versa*. The anterior end is regular, rounded and broad while the posterior end is usually slightly bent (Fig. 1 a) or hook-like (Fig. 1 b). In some cases both the extremities may be

recurved and afflicted or the parasite may show a constriction about its centre. These elongate forms measure $14.4 \mu\text{m} \times 2.95 \mu\text{m}$ with an average area of $28.95 \mu\text{m}^2$ occupying about 25.97% of the total host cell-parasite complex. Granulated cytoplasm stains azure blue or faint pink with Giemsa stain. The nucleus, approximately central in position, almost fills up the entire width of the parasite. It may be rounded (Fig. 1 f), oval (Fig. 1a) or band-shaped or irregular in outline, measuring $3.25 \mu\text{m} \times 1.95 \mu\text{m}$ and $5.18 \mu\text{m}^2$ in area ; stains pink with Giemsa. The

gametocytes are always enveloped in a pink coloured thick capsule.

Intermediate and young forms (Fig. 1 c) :

Intermediate forms measure $10.5 \mu\text{m} \times 3.0 \mu\text{m}$ and are comparatively rare in the peripheral circulation. Cytoplasm stains light blue and contains a few metachromatic granules. The nucleus is the aggregation of chromatin materials at the centre of the parasite. Young forms measure $5.2 \mu\text{m} \times 2.5 \mu\text{m}$; both the ends rounded, capsule-like; band-shaped nucleus at the centre. Cytoplasm stains deep blue, with very few metachromatic granules.

Extracorporeal free forms (Fig. 1 d & Pl. V, Fig. 5) :

These are elongated with one end rounded and the opposite end being attenuated; measuring $21.72 \mu\text{m} \times 2.84 \mu\text{m}$ with $24.8 \mu\text{m}^2$ in area. The cytoplasm is bluish with metachromatic granules evenly distributed. The compact rounded nucleus measures $3.2 \mu\text{m} \times 2.28 \mu\text{m}$ with an area of $4.9 \mu\text{m}^2$.

Effect of the parasite on the host cell :

Elongate forms caused a marked hypertrophy of the infected red blood corpuscles and their nuclei. As a result of which the host cell became enlarged and the host cell nucleus was placed peripherally or sometimes pushed to apical position in case of double infection (Fig. 6). Nuclear Displacement Ratio=0.17. Host cell hypertrophied in length (7.3%) and area (11.3%). Host cell nucleus also hypertrophied in area (17%).

Uninfected erythrocyte :

N=15. Cell $15.47 \mu\text{m} \times 8.55 \mu\text{m}$ and $100.10 \mu\text{m}^2$ in area. Cell nucleus $5.85 \mu\text{m} \times 2.72 \mu\text{m}$ and $13.28 \mu\text{m}^2$ in area. N. D. R.=0.97.

Schizogony (Pl. V, Figs. 9-12) :

A thorough examination revealed some schizogonic stages in the liver parenchymatic cells. Erythrocytic schizogony was not detected. The early developmental stage of a schizont is an oblong or spherical uninucleate body measuring $4.5 \times 4.2 \mu\text{m}$; occupying a cavity (Parasitophorous vacuole) (Fig. 9) within the liver cell. The spherical nucleus is $2.5 \mu\text{m}$ in diameter, surrounded by an areola. The multinucleate schizonts (Fig. 11) measure $15.00 \mu\text{m} \times 14.00 \mu\text{m}$. The larger mature schizonts measure $20 \mu\text{m} \times 13 \mu\text{m}$ and contain many slender curved merozoites ($4.5 \mu\text{m} \times 1.5 \mu\text{m}$).

A more or less spherical nucleus is situated at the centre of each merozoite which stains deep black with Iron alum haematoxylin stain.

DISCUSSION

Haemogregarina nucleobisecans along with its hepatic schizogony was first reported by Shortt (1917) from North India. Wenyon (1926) reported the same parasite as *Haemogregarina bisecans* from the same anuran host of India.

The present study of *H. nucleobisecans* well agrees with that of Shortt's description in all general respects. The bisecting nature of the host-cell nucleus, however, has not been observed by the present author. Shortt (1917) did not observe any movement of this parasite but the present author noted a characteristic slow movement of extracorporeal gametocytes in the citrated saline preparation of fresh blood.

H. nucleobisecans has also been recorded during the present investigation from *Bufo andersoni* which is little smaller than that described from the type host. Moreover, in *B. andersoni* some leucocytes were also found

to be infected with the parasites measuring $11.5 \mu\text{m} \times 2.5 \mu\text{m}$.

Tissue Schizogony of the parasite has been observed in both the hosts *Bufo melanostictus* and *B. andersoni*. One type of schizogony with only one type of merozoite was noted.

Haemogregarina nucleobisecans has been recorded from a new geographical locale of Eastern India and from a new host *Bufo andersoni* along with *B. melanostictus*.

ACKNOWLEDGEMENT

The author is grateful to Dr. B. K. Tikader, Director, Zoological Survey of India for providing laboratory facilities. He is also indebted to Prof. A. Choudhury, Calcutta University and Dr. A. K. Mandal, Superintending Zoologist, Zoological Survey of India for their interest in the study.

REFERENCES

- PEARSE, A. G. E. 1960. *Histochemistry*. 2nd Edn. J. and A. Churchill, London.
- RAY, R. 1977. A note on the life-cycle of a *Haemogregarina* sp. from an Indian Ranid frog. *Proc. 1st. Natl. Cong. Parasitology*, Baroda, India pp. 16-17 (Abst.).
- RAY, R. 1979. 'Studies on the Haematozoa of Indian Amphibians'—Ph. D. Thesis, Calcutta University, 354 pp.
- SHORTT, H. E. 1917. Notes on two Haemogregarines of cold-blooded vertebrates. *Indian J. med. Res.*, 4 : 402-413.
- SINHA, C. K. 1979. A haemogregarine from a Himalayan toad, *Bufo himalayanus* Günther. *Proc. 2nd Natl. Cong. Parasitology*, B. H. U. Varansi, India. pp. 58 (Sec-I) (Abst.).
- WENYON, C. M. 1926. *Protozoology*. 2 vols. 1563 pp. Bailliere, Tindall and Cox, London.